

Developed Galantamine Therapy for Alzheimer's Disease by Introducing Nano-Drug Delivery Systems

Walaa Ahmed Mostapha¹ and Shewikar Tewfik El-Bakry^{2,*}

¹Zoology Department, Women's College for Arts, Science and Education – Ain Shams University, Egypt

²Psychiatry Department, Faculty of Medicine, Benha University, Egypt

Abstract: The major cause of dementia, a major public health problem, is Alzheimer's disease (AD). A reliable method for the diagnosis and follow-up of Alzheimer's disease is needed together with a specific biological marker. Galantamine, an acetylcholinesterase inhibitor for AD therapy has several reported side effects. One approach to reduce dosing amounts, frequency of administration, and adverse side effects while maintaining the drug efficiency, is the development of drug delivery systems using nanoparticles. Presently Galantamine (Gal) coated with either Cerium/ Calcium hydroxyapatite (Ce/Ca-HAp) or carboxymethyl chitosan/ Ceria/ Calcium hydroxyapatite (CMCS/Ce/Ca-HAp) was *i.p.* injected at a dose of 2.5 mg/kg b.wt. for 2 and 4 weeks. 86 female adult albino Wistar rats (189- 200 gm weight) were used. Alzheimer was induced in ovariectomized rats by Aluminium chloride (AlCl₃) oral treatment at doses of 17mg/kg b.wt. daily for 2 months. Rats were divided into six groups: Group (1) normal control; Group (2) Galantamine *i.p.* injected at a dose of 2.5 mg/kg b.wt; Group (3) Alzheimer ; Group (4) Alzheimer's induced rats treated *i.p.* with Gal; Group (5) Alzheimer's induced rats treated with Gal. coated with Ce/Ca-HAp; Group (6) Alzheimer's induced rats treated with Gal. coated with Gal coated with CMCS/Ce/Ca-HAp for 2 and 4 weeks. In the concurrent study, AD induced histological alterations manifested as amyloid plaque formation of different sizes; congestion with perivascular edema; degenerated neurons with diffused gliosis; loss of pyramidal cells; separation of cortical tissue and formation of fibrous glial scar. Several tests may be cumulatively used for early detection as decreased Ach; Bcl2; Tpl; GSH; SOD; CAT; Cyto P450 and increased Ab; Chol; B-FABP; NO; MDA; GSSH. Treatment with Gal coated by Ce/Ca-HAp imposed highly significant improvement to near to normal levels in both histological and biochemical parameters. Gal coated by CMCS/Ce/Ca-HAp failed to encounter obvious ameliorations. In conclusion, brain markers (Ach; Bcl-2; A β ; TPL; Chol; B-FABP; NO; MDA) together with brain antioxidants (GSH; SOD; CAT; CytoP450) may impose progressive laboratory testing method besides well known imaging techniques. Galantamine therapy may impose limited improvements thus drug delivery systems Gal coated by Ce/Ca-HAp may aid to minimize dosing amounts, frequency of administration, and adverse side effects of drug while increasing its therapeutic efficacy.

Keywords: Alzheimer (Alz), Ceria-doped calcium hydroxyapatite (Ce/Ca-HAp), Carboxymethyl Chitosan/Ceria/hydroxyapatite composite (CMCS/Ce/Ca-HAp), Galantamine, Rat, Histology and Biochemistry.

INTRODUCTION

Dementia is one of the major public health problems, where increasing numbers of patients impose a major financial burden on health care systems. It is majorly related to aging, and with the increasing numbers of elderly people in the population, the number of patients with dementia is growing rapidly. The major cause of dementia is Alzheimer's disease (AD). It is a progressive neurodegenerative disorder of the brain that is characterized by loss of neurons because of extracellular accumulation of amyloid beta (Ab) and intracellular hyperphosphorylation of the tau protein [1].

Risk factors for AD are age and a positive family history of dementia, since more than one-third of AD patients have one or more affected first degree relatives [2]. Other risk factors that may be associated with the development of AD include severe head trauma, low levels of education, female gender, previous depression, and vascular factors [3, 4].

Neuropsychological tests are often influenced by education [5], practice [6], and sociocultural or ethnic factors [7]. As a screening test for dementia, the sensitivity of MMSE (mini-mental state examination) test has been 56-90 % and specificity has varied from 85 % to 95 % [8].

The two major microscopic lesions for AD are amyloid plaques and neurofibrillary tangles (NFT), which are found significantly more often in AD compared to normal aging. Computed tomography (CT) does not differentiate early AD from normal aging with high diagnostic accuracy [9]. Single photon emission computed tomography (SPECT) may be useful in the differential diagnosis of dementia [10]. Magnetic resonance imaging (MRI) offers a superior anatomic discrimination power and permits accurate imaging of the affected regions.

Thus, a reliable method for the diagnosis and follow-up of AD is needed.

An ideal biomarker should detect a fundamental feature of the neuropathology of AD, be able to detect AD early in the course of the disease with high

*Address correspondence to this author at the Psychiatry Department, Faculty of Medicine, Benha University, Egypt; Tel: 01005502606; E-mail: shewikare@yahoo.com

sensitivity, distinguish it from other dementias, and be validated in neuropathologically confirmed AD cases.

A biomarker reflecting neuropathological changes at the molecular level in AD brain would be very useful in the differential diagnosis of dementia, and in distinguishing AD patients from those individuals with mild cognitive impairment who do not develop AD, and from patients with depression [11].

Studies on biochemical markers for the diagnosis of AD in cerebrospinal fluid (CSF) and blood are based on the detection of inflammatory proteins, markers of cholesterol homeostasis, oxidative stress, or related to characteristic pathological alterations in AD [12].

Cognitive deficits in AD have been widely associated with dysfunction of the cholinergic system [13] consequent upon degeneration of cholinergic cell bodies in the basal forebrain [14]. In post-mortem frontal cortex from AD patients, acetylcholine (ACh) levels, cholinacetyltransferase (ChAT) and acetylcholinesterase (AChE) activity were all reduced compared to controls [15]. Consequent to the cholinergic hypothesis of dementia, animal studies have been used to evaluate the role of cholinergic neurotransmission in learning and memory and selective lesion of cholinergic neurones remains nowadays as a widely used method to mimic some aspects of neurodegeneration in AD [16].

Bcl-2 (B-cell lymphoma 2), is a regulator protein that regulates cell death (apoptosis), by either inducing (pro-apoptotic) or inhibiting (anti-apoptotic) apoptosis. That Bcl-2 is expressed in neurons in the aged human brain complements observations suggesting neuronal death by apoptosis in AD patients [17]. A critical role of the β -amyloid ($A\beta$) in the pathogenesis of AD has been supported by human, animal, and *in vitro* studies [18]. Intracellular and membrane-associated $A\beta$, especially $A\beta$ 42 in the temporal neocortex, may be more closely related to AD symptoms than other measured $A\beta$ species [19].

Tissue factor thromboplastin TPL is the primary initiator of the extrinsic coagulation pathway. In contrast to other coagulation factors, it is an integral membrane glycoprotein that does not circulate as a plasma protein [20]. Although tissue factor might potentially contribute to the formation of senile plaques, it could also accumulate in the plaques as a secondary response to other biochemical perturbations [21].

Cholesterol plays an important role in promoting the deposition of amyloid in the brain [22]. They reported

that participants who had higher levels of LDL cholesterol and lower levels HDL cholesterol had higher levels of amyloid in the brain. Brain-derived metabolites of cholesterol also appear to be elevated in the early stages of disease prior to the onset of cognitive impairment rendering them an important marker of underlying cerebrovascular disease preceding cognitive impairment and risk for developing cognitive impairment [23].

Brain-type fatty acid binding proteins (B-FABP) [24] may be suitable markers for the detection of brain injury. B-FABP is a member of a family of nine distinct FABP types, each named after the tissue in which it was first detected [25]. They are rapidly released from damaged cells into the circulation and are cleared from the circulation by the kidney with a plasma half-life of 20 min [26]. In adult-stage mice, B-FABP is produced in very low concentrations and is detected only in glial cells (presumptive astrocytes) of the white matter [27].

Nitric oxide NO is an atypical neurotransmitter formed from L-arginine by the enzyme nitric oxide synthase (NOS) [28]. NO is a well-known physiological signaling agent, and a pleiotropic regulator in various pathologies including tumor growth and AD [29]. (NO-) dependent oxidative stress results in mitochondrial ultrastructural alterations and DNA damage in cases of AD [30]. In fact, it plays a crucial role in mitochondrial respiration [31], since even low (nanomolar) concentrations of NO were found to reversibly inhibit the mitochondrial respiratory chain enzyme cytochrome oxidase (complex IV) and compete with molecular oxygen.

MDA arises largely from the peroxidation of PUFA. A number of studies document elevated levels of MDA in AD in the plasma/serum [32, 33]. The most prevalent antioxidant in most brain cells is GSH. It can react with ROS and oxidized products forming glutathione disulphide (GSSG), either catalysed by Glutathione Peroxidase (GPx) or independently. The GSSG can then be converted back to reduced GSH by Glutathione Reductase (GR). Studies of human brains have indicated that the ratio between reduced and oxidized glutathione (GSH/GSSG) is decreased in affected brain regions of AD patients compared to controls [34]. Superoxide dismutase (SOD) is part of the initial defense against ROS and catalyses the conversion of O_2^- to H_2O_2 and oxygen (O_2). The activity of SOD in serum was reduced in AD patients compared to controls and was negatively correlated to the lipid peroxidation marker MDA [35].

Galantamine, is an acetylcholinesterase inhibitor that may serve to prevent hypertension from precipitating vascular dementia, which is the second most common cause for dementia after AD. The cerebrovascular injury caused by hypertension has been associated with compromised cognitive function, mainly memory dysfunction and decreased speed of information processing. It was reported that galantamine works by enhancing cholinergic function by increasing the concentration of acetylcholine in the brain and enhancing cholinergic neuro-transmission in the brain [36]. Yet side effects from the drug have been thoroughly reported where higher doses are gradually required.

One approach to reduce dosing amounts, frequency of administration, and adverse side effects while maintaining the drug efficiency, is the development of new drug delivery systems with inflammatory site targeting and long circulating time [37].

Hydroxyapatite (HAp) has been extensively used in medicine for implant fabrication owing to its similarity with mineral constituents found in hard tissues [38], which leads to formation of bonds between the bone and the implanted materials being acted as biocompatible phase reinforcement in composites as well coatings to metal implants and granular fillers for direct incorporation into human tissues. Implementation of new species in the HAp lattice offer fundamentally new possibilities and areas of their practical applications in biology and medicine

Again Ceria (CeO₂) nanoparticles exhibit high catalytic activity and a regenerative capacity to neutralize ROS. Ceria was found to protect cells against oxidative stress, inflammation, or damage caused by radiation. The particles are small and can cross the blood brain barrier [39].

Carboxymethyl chitosan (CMCS) is one of the most investigated water-soluble derivatives of chitosan (CS) that own specific biological activities such as antitumor activity, immune-stimulating effects, enhancing protective effects against infection with some pathogens in mice, antimicrobial activity and radical scavenging activity. Because of the carboxymethylation, CMCS possesses negative charges when dissolved in water, the CMCS hydrogels seem to be successfully prepared by physical crosslink with calcium-based biopolymers [40].

Successful therapeutic regimens for incurring favourable pathological influence for AD necessitate

reliable earlier diagnosis for the onset of clinical settings. They should be highly specific to AD especially in the early stages of the disease [1].

Thus the aim of the presented study was to identify several sensitive biological markers to detect fundamental features of the neuropathology of Alzheimer's disease in its earlier stages. As Galantamine seems to impose serious side effects besides its poor therapeutic capabilities new delivery systems using nanoparticles are given.

MATERIAL AND METHODS

86 female adult albino Wistar rats (189- 200 gm weight) from the Animal breeding colony of the Medical Research Centre Ain Shams University were presently used. After a one week pre-experimentation adaptation (food and water ad libitum) period animals were ovariectomized surgically as several lines of evidence suggest that changes in hormones after menopause may play an important role in the incidence of cognitive dysfunction & also in the development of the AD [41]. Alzheimer induced rats were ovariectomized prior to Aluminium chloride (AlCl₃) oral treatment at doses of 17mg/kg b.wt. [42] daily for 2 months after one month post-operative procedure.

Galantamine was *i.p.* injected at a dose of 2.5 mg/kg b.wt. for 2 and 4 weeks [43].

Galantamine was coated with either Cerium/ Calcium hydroxyapatite (Ce/Ca-HAp) or carboxymethyl chitosan/ Ceria/ Calcium hydroxyapatite (CMCS/Ce/Ca-HAP) and *i.p.* injected at a dose of 2.5 mg/kg b.wt. for 2 and 4 weeks.

Animals were allotted as follows:-

- a) *Group 1 control (C)*: Gonad intact animals (8)
- b) *Group 2 Galantamine (Gal)*: Gonad intact animals (8) treated with galantamine.
- c) *Group 3 Alzheimer (Alz)*: Ovariectomized animals (22) treated orally with AlCl₃ (17 mg/kg b.wt) daily for 2 months following one month surgery.
- d) *Group 4 (Alz + Gal)*: Alzheimer's disease-induced rats (16) treated *i.p.* with Gal (2.5 mg/kg b.wt.).
- e) *Group 5 (Alz + Gal. + Ce/Ca-HAp)*: Alzheimer's disease-induced animals (16) treated (*i.p.*) with Gal. coated with Ce/Ca-HAp (2.5 mg/kg b.wt.).

- f) *Group 6 (Alz +Gal. + CMCS/Ce/Ca-HAP):* Alzheimer's disease-induced animals (16) treated *i.p* with Gal coated with CMCS/Ce/Ca-HAp (2.5 mg/kg b.wt.).

After 2 and 4 weeks rats were ether inhalation anaesthetized where brains were collected from all groups. One half of each brain was homogenized immediately to give 10% (w/v) homogenate in ice-cold medium containing 50 mM Tris-HCl (pH 7.4) and 300 mM sucrose. The homogenate was centrifuged at 3000 rpm for 10 min at 4 °C. The supernatant (10%) was separated for biochemical estimations.

The quantitative measurement of brain markers included Ach levels [44]; Bcl-2, [45]; A β [46]; TPL [47]; Chol [48]; B-FABP [49]; NO [50] and MDA [51]. Brain antioxidants included GSH [52], GSSH, SOD [53], CAT [54] and CytoP₄₅₀ reductase [55].

The other half was washed in saline (0.9% NaCl) and placed accordingly in 10% neutral buffered formalin for fixation and processed for histological analysis by Haematoxylin and Eosin for general histological examination [56].

RESULTS

Histological Investigations

Hx&E staining data are shown in Figure 1 in control (Figure 1A) and experimental groups (Figure 1B-1H). Sections of rat brain from Gal animals manifested no alterations from control (1B). On the other hand following Alz induction, amyloid plaques of different sizes, congestion with perivascular edema, degenerated neurons with diffused gliosis, loss of pyramidal cells, separation of cortical tissue and formation of fibrous glial scar were manifested (Figure 1C & 1D). Following Gal therapy rat brain sections presented mild attenuation in amyloid plaques which persisted in a less aggressive manner with the progress of time; peri-vascular edema accompanied by diffused gliosis and degeneration in some of the hippocampal neurons (Figure 1E). Using Gal coated by Ce/Ca-HAp designated near to normal patterns following 4 weeks of treatment (Figure 1F). While Gal coated by CMCS/Ce/Ca-HAp failed to induce ameliorative levels of recovery (Figure 1G & 1H).

Biochemical Investigations

Brain Marker Profile

Concurrent results signify significant decreases in Ach, Bcl-2 and TPL in brain tissue from Alz-induced

rats as compared to normal control animals. On the contrary, significant increases in Ab, Chol, BFABP NO and MDA were recorded in the same group of rats. All parameters were significantly ameliorated to near to normal figures following treatment with Gal coated by Ce/Ca-HAp and not Gal coated by CMCS/Ce/Ca-HAp which failed to impose ameliorative figures (Table 1).

Brain Enzymes Antioxidants Profile

Presently, significant depletion was obvious in GSH, SOD, CAT and CytoP₄₅₀ reductase in brain tissue of Alz-induced rats as compared with normal control. Partial amendment in above parameters was reached following Galantamine treatment. Treatment with Gal coated by Ce/Ca-HAp imposed highly significant improvement to near to normal levels in the previous parameters. On the contrary, Gal coated by CMCS/Ce/Ca-HAp failed to encounter any obvious ameliorations. These results validate fore mentioned histopathological alterations (Tables 1 & 2).

DISCUSSION

Alzheimer's disease (AD) is a progressive neurodegenerative disorder that is the most common form of dementia, accounting for approximately 50-60% of all cases [57]. The disorder usually starts in a mild form and progressively gets worse because of late discovery of onset together with the lack of specific efficient therapeutic regimens and due to its progressive course.

Many studies of putative biomarkers have been conducted, but none of the antemortem tests has fulfilled all the criteria for an ideal biomarker. This necessitates the use of several methodology for the early identification together with the different diagnostic imaging methods as photon emission computed tomography (SPECT) and Magnetic resonance imaging (MRI)). Such biomarkers include Ach, Bcl₂, amyloid peptide, TPL, Chol, BFABP, NO and MDA in addition to the brain antioxidant enzymes GSH, GSSH, SOD, CAT and CyoP₄₅₀. Biomarkers seem to be more sensitive to Alzheimer changes in its earlier stages as presently designated that would impose a reliable method for early diagnosis.

In the concurrent study, AD was reported to induce serious histological alterations manifested as amyloid plaque formation of different sizes; congestion with perivascular edema; degenerated neurons with diffused gliosis; loss of pyramidal cells; separation of cortical tissue and formation of fibrous glial scar.

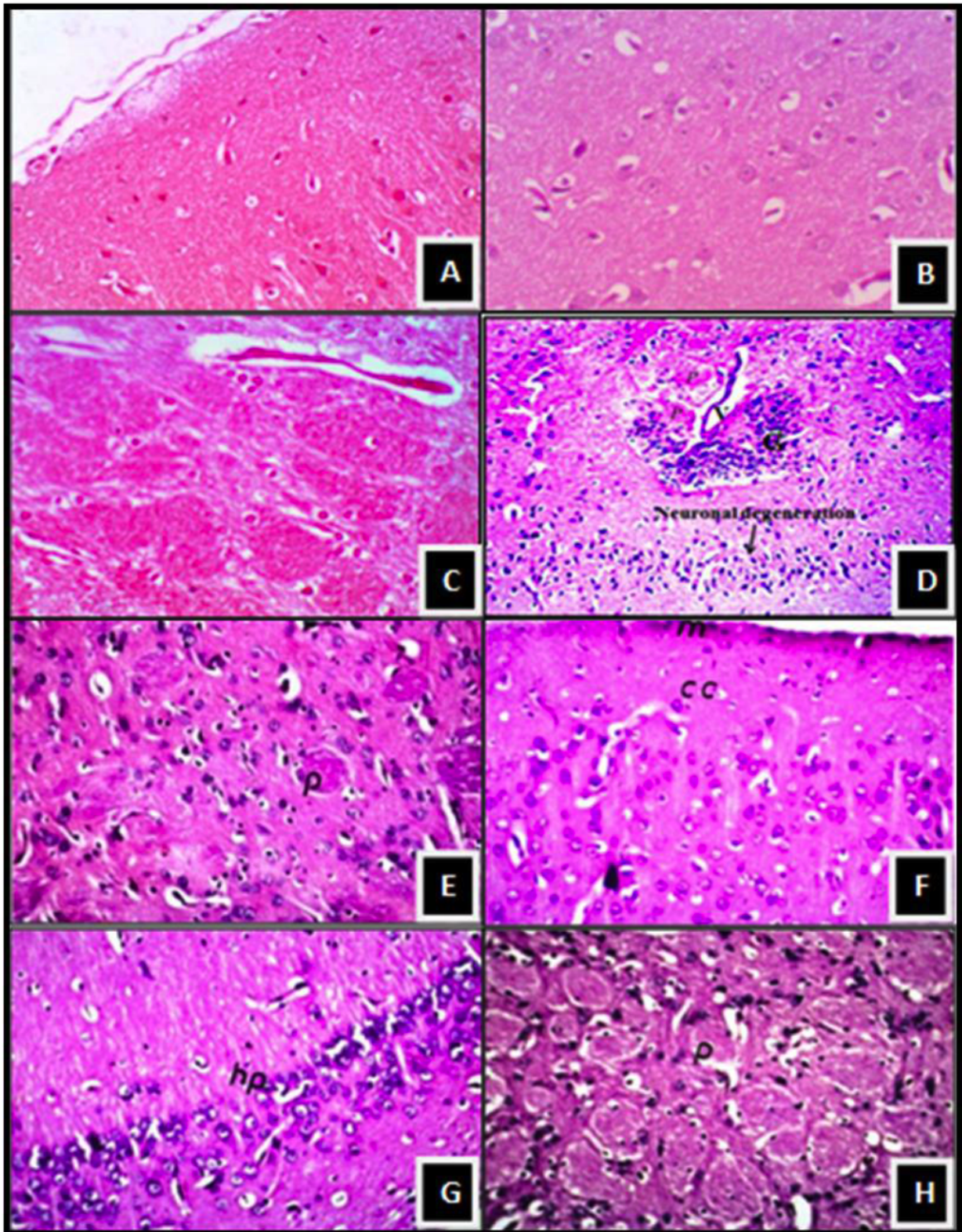


Figure 1: Photomicrograph of brain sections of rats from control and experimental groups. (A): normal control. (B): Gal treated showing no alterations from control. (C & D): Alz induced showing amyloid plaques of different sizes, perivascular edema and diffused gliosis. (E): Alz induced after Gal therapy showing persistence of amyloid plaques and diffused gliosis. (F & G): Alz induced treated with Gal coated by Ce/Ca-HAp showing near to normal patterns in cerebral cortex and hippocampus. (H): Alz induced treated with Gal coated by CMCS/Ce/Ca-HAp showing persistence of lesions. (H E; X 200) (p) amyloid plaques; (cc)cerebral cortex; (hp) hippocampus.

Table 1: Concentration of Different Brain Markers in Brain Tissue Samples from Alz-Induced and Treated Rats with Galantamine, Gal+Ce/Ca-HAp and Gal+CMCS/Ce/Ca-HAp

		Control	ALZ	ALZ+ Gal	Alz+Gal+Ce/ Ca-HAp	Alz+Gal+CMCS /Ce/Ca-HAp
Ach(μ mol/ mg protein)	2 w	100.67 ^A _a ± 3.22	71.68 ^B _a ± 2.24	83.29 ^C _a ± 2.84	90.54 ^D _a ± 2.93	84.01 ^E _a ± 2.87
	4 w	101.42 ^A _a ± 3.32	67.55 ^B _b ± 2.16	94.68 ^C _b ± 3.09	97.76 ^D _b ± 3.23	94.88 ^E _b ± 3.11
Bcl-2 (pg/mg protein)	2 w	92.33 ^A _a ± 2.91	58.93 ^B _a ± 1.78	71.45 ^C _a ± 2.31	77.31 ^D _a ± 2.47	72.04 ^E _a ± 2.35
	4 w	92.42 ^A _a ± 2.94	54.27 ^B _b ± 1.49	80.73 ^C _b ± 2.55	86.55 ^D _b ± 2.69	81.26 ^E _b ± 2.58
Amyloid peptide (pg/mg protein)	2 w	13.26 ^A _a ± 0.53	25.77 ^B _a ± 0.93	19.05 ^C _a ± 0.76	17.25 ^D _a ± 0.69	18.79 ^E _a ± 0.71
	4 w	13.21 ^A _a ± 0.51	29.02 ^B _b ± 1.12	15.82 ^C _b ± 0.62	14.44 ^D _b ± 0.58	15.63 ^E _b ± 0.64
TPL (μ g/mg protein)	2 w	1277.4 ± 21.59	785.41 ± 15.78	953.68 ± 19.25	1077.43 ± 21.03	961.22 ± 19.43
	4 w	1259.8 ± 20.87	694.33 ± 14.25	1104.55 ± 22.17	1269.55 ± 22.86	1127.38 ± 22.06
Chol (μ g/mg protein)	2 w	648.66 ^A _a ± 13.16	1008.29 ^B _a ± 23.56	870.56 ^C _a ± 18.82	801.21 ^D _a ± 17.41	863.98 ^E _a ± 18.77
	4 w	652.51 ^A _a ± 13.19	1152.65 ^B _b ± 25.78	749.91 ^C _b ± 15.33	682.32 ^D _b ± 14.62	736.72 ^E _b ± 15.18
B-FABP (pg/mg protein)	2 w	39.91 ^A _a ± 2.14	75.99 ^B _a ± 3.86	58.54 ^C _a ± 2.78	53.76 ^D _a ± 2.52	57.88 ^E _a ± 2.69
	4 w	40.52 ^A _a ± 2.23	81.31 ^B _b ± 4.09	50.27 ^C _b ± 2.39	46.56 ^D _b ± 2.28	49.55 ^E _b ± 2.37
NO (pg/mg protein)	2 w	36.43 ^A _a ± 1.97	70.67 ^B _a ± 2.94	51.31 ^C _a ± 2.28	47.22 ^D _a ± 2.11	50.82 ^E _a ± 2.23
	4 w	35.99 ^A _a ± 1.95	76.55 ^B _b ± 3.18	43.77 ^C _b ± 2.04	39.81 ^D _b ± 1.96	43.49 ^E _b ± 2.02
MDA (nmol/mg protein)	2 w	4.07 ^A _a ± 0.101	8.02 ^B _a ± 0.206	6.28 ^C _a ± 0.154	5.79 ^D _a ± 0.146	6.08 ^E _a ± 0.151
	4 w	4.13 ^A _a ± 0.103	8.61 ^B _b ± 0.273	5.51 ^C _b ± 0.138	5.05 ^D _b ± 0.123	5.39 ^E _b ± 0.134

Values are expressed as mean ± SE.

^{A,B,C,D}Means with a common superscript within a row are statically significantly different (P<0.05).

^{a,b}Means with a common subscript within a column are statically significantly different (P<0.05).

Table 2: Concentration of Different Brain Enzyme Antioxidants in Brain Tissue Samples from Alz-Induced and Treated Rats with Galantamine, Gal+Ce/Ca-HAp and Gal+CMCS/Ce/Ca-HAp

		Control	ALZ	ALZ+ Gal	Alz+Gal+Ce/ Ca- HAp	Alz+Gal+CMCS /Ce/Ca-HAp
GSH (U/mg protein)	2 w	41.17 ^A _a ± 1.423	26.89 ^B _a ± 0.832	33.26 ^C _a ± 0.957	35.07 ^D _a ± 0.988	33.79 ^E _a ± 0.972
	4 w	40.93 ^A _a ± 1.418	23.47 ^B _b ± 0.795	37.91 ^C _b ± 1.086	40.54 ^D _b ± 1.302	38.11 ^E _b ± 1.117
GSSG (μ M/mg protein)	2 w	0.63 ^A _a ± 0.008	1.16 ^B _a ± 0.018	0.82 ^C _a ± 0.012	0.74 ^D _a ± 0.011	0.81 ^E _a ± 0.013
	4 w	0.64 ^A _a ± 0.008	1.42 ^B _b ± 0.024	0.74 ^C _b ± 0.011	0.65 ^D _b ± 0.009	0.74 ^E _b ± 0.012
SOD (U/mg protein)	2 w	3.08 ^A _a ± 0.071	1.69 ^B _a ± 0.034	2.55 ^C _a ± 0.045	2.79 ^D _a ± 0.052	2.55 ^E _a ± 0.045
	4 w	2.9 ^A _a ± 0.068	1.48 ^B _b ± 0.029	2.87 ^C _b ± 0.057	2.95 ^D _b ± 0.071	2.91 ^E _b ± 0.069
CAT (U/mg protein)	2 w	6.50 ^A _a ± 0.173	5.09 ^B _a ± 0.101	5.78 ^C _a ± 0.136	6.08 ^D _a ± 0.148	5.69 ^E _a ± 0.133
	4 w	6.49 ^A _a ± 0.168	4.78 ^B _b ± 0.092	6.03 ^C _b ± 0.149	6.43 ^D _b ± 0.171	5.97 ^E _b ± 0.152
Cyto P ₄₅₀ reductase (ng/mg protein)	2 w	2.19 ^A _a ± 0.023	1.23 ^B _a ± 0.017	1.82 ^C _a ± 0.021	2.11 ^D _a ± 0.029	1.96 ^E _a ± 0.025
	4 w	2.11 ^A _a ± 0.024	0.958 ^B _b ± 0.012	2.06 ^C _b ± 0.024	2.34 ^D _b ± 0.034	2.16 ^E _b ± 0.028

Values are expressed as mean ± SE.

^{A,B,C,D}Means with a common superscript within a row are statically significantly different (P<0.05).

^{a,b}Means with a common subscript within a column are statically significantly different (P<0.05).

Similarly, neuropathological examination of the AD rat brain showed extensive neuronal loss, accumulation of fibrillary amyloid β plaques, and neurofibrillary tangles within neurons [58]. Mahdy *et al.* [59] and Yassin *et al.* [60] observed necrosis of the brain, spongy

appearance, plaques, and loss of normal structure, various sizes of amyloid plaques in the hippocampus, neurofibrillary tangles and fatty changes in Alzheimer induced rat brain tissue.

Aluminum salt has been found to induce the overexpression of APP. Present manifestations may be primarily a sequel to inflammatory responses that are known to play an important role in neurodegenerative disease. Vallés *et al.* [61] demonstrated that A β peptide causes oxidative stress in the neurons and inflammation in the astrocytes indicating that this toxic peptide can affect not only neuronal cells but astrocytes too. Wu *et al.* [62] also suggested that, the amyloid plaques caused by AI administration plays a role in the pathology of AD by directly inducing neuronal cytotoxicity and stimulating microglia to secrete cytokines and reactive oxygen species (ROS) which also damage neurons. Amyloid plaques could activate astrocytes and oligodendrocytes to produce chemokines, in particular Monocytes chemotactic protein (MCP-1), which serves as potent *in vitro* microglial and macrophage chemoattractants.

The challenges for establishing an early diagnosis of Alzheimer's disease (AD) have created a need for biomarkers that reflect the core pathology of the disease making earlier diagnosis possible in a clinical setting. They should be highly specific to AD and sensitive to changes, especially in the early stages of the disease [1].

Recorded results signify a considerable decrease in Ach content and a significant increase in Bcl-2 and TPL in Alzheimer induced rats. Contrary to that a limited increase in Ab was recorded and a significant increase in Chol, BFABP, NO and MDA.

Cognitive deficits in neuropsychiatric disorders have been closely related to cholinergic deficits where total Ach levels in both human and rats were reduced [15]. The cholinergic hypothesis suggests that a dysfunction of acetylcholine containing neurons in the brain contributes substantially to the cognitive decline observed in those with advanced age and AD [63].

Decreased expression of bcl-2 is observed in Alzheimer's dementia in neurons with neurofibrillary tangles [64]. Studies undertaken from Bcl-2 localization (mitochondria and endoplasmic reticulum) suggest that it inhibits generation of reactive oxygen species or lipid peroxidation [65]. It is also reported to modulate calcium fluxes from intracellular stores in hippocampal neurons *in vitro* [66].

Overproduction of amyloidogenic forms of the amyloid-(A β), a peptide that is generated by a sequential cleavage of APP by the β - and γ -secretases

[67] is the hallmark of AD. Similarly, present studies have signified significant increase in A β in AD rats. A β is a complex biological molecule which interacts with many types of receptors and/or forms insoluble assemblies and, eventually, its nonphysiological depositions alternate with the normal neuronal conditions [68]. In aging and pathological conditions, the formation and clearance of A β are disturbed leading to an accumulation of A β and senile plaque formation. Neurodegeneration in AD is mediated in part through soluble forms of A β . Increased soluble A β concentration correlates with cognitive decline in AD-affected individuals. Hence, it may be considered as a reliable predictor of AD [68].

TPL is an integral membrane protein that is normally sequestered from circulating factor VII and other coagulation factors. Tissue injury exposes TPL to circulating factor VII resulting in complex formation and initiation of blotting cascade via the extrinsic pathway [21] In the concurrent study marked decrease in TPL has been recorded.

It has been suggested that increased levels of free cholesterol in neuronal cell membranes may provoke A β formation [60]. Highly significant augmented levels for cholesterol have been presently well documented. Several related studies have reported concomitant findings as Hughes [23] and Reed *et al.* [22]. Experimental *in vitro* and *in vivo* data indicate that brain cholesterol homeostasis is coupled with brain amyloid metabolism [70]. Cell culture studies demonstrate that membrane cholesterol controls the direction of the processing of the APP *in vitro* [70]. Again, under *in vitro* conditions, cholesterol has been shown to influence a number of processes involved in the generation of A β peptide of residues and neurofibrillary tangles [71].

In a study of patients B-FABP levels were elevated in serum of 29% of the patients with AD. B-FABP expression was observed in reactive astrocytes in brain sections. Thus it could be used in an attempt to reveal a sensitive marker for various neurodegenerative diseases and can therefore have importance for defining subgroups of these patients [72].

NO (produced by endothelial cells of blood vessels) is a protective molecule that maintains immune, cardiovascular, nervous, kidney, stomach and intestinal, skin and other beneficial effects. Accordingly, lack of NO production by endothelial cells due to increase in amyloid beta peptide aggregate formation and/or free radical generation only contributes to

pathology of AD. It was verified by [73] that A β produced nitric oxide which resulted in a change to the protein Drp1, called SNO-Drp1. This change, which was the attachment of NO to the protein, assisted by cysteine at the 644th position, caused an increase in mitochondrial fragmentation and cell death in Alzheimer's brains [74].

Increased levels of MDA in the brain in AD have been confirmed by several studies [1]. One of the main reasons for high malondialdehyde (lipid peroxidation) in elder patients could be melatonin deficiency as decrease in melatonin seems to be related to aging. The authors emphasized that low melatonin levels could be explained not only by a decrease in melatonin due to aging, but also by a decrease in melatonin due to AD.

Oxidative stress is a significant element in AD pathogenesis. Possibly, lack of protection against ROS production in the aging brain could be one triggering cause of AD. Oxidative stress is one of the first consequences of A β overproduction in the brain. The pressure from oxidative stress in the aging brain in combination with a lowered antioxidant defense creates a harmful combination that could disturb functions and damage organelles such as mitochondria. This would eventually lead to loss of synapses and cell death that give rise to the clinical symptoms associated with AD [75].

The activity of SOD is a sensitive index in oxidative damage as it scavenges the superoxide anion to form hydrogen peroxide leading to diminished toxic effects. Also, Glutathione reduced, glutathione reductase and glutathione-S-transferase are closely related to the direct elimination of reactive oxygen species. Therefore, the reduction in the activity of these enzymes may result in a number of deleterious effects due to the accumulation of superoxide radicals and hydrogen peroxide, linked with neurodegenerative diseases [76]

Galantamine the acetylcholinesterase inhibitor, has long been the drug of choice for treatment of AD patients for its recorded beneficial role in the slowing down of the degenerative outcomes on the brain that cause impaired memory. Pathways of Galantamine therapy is through preventing acetylcholinesterase from breaking down acetylcholine in the brain. As a result, an increased concentration of acetylcholine leads to increased communication between nerve cells. This may temporarily alleviate or stabilize some symptoms

of Alzheimer's disease. In the present study therapeutic doses of Galantamine to AD rats signified limited ameliorations in the studied parameters of brain markers and antioxidant. Similarly, Galantamine coated by CMCS/Ce/Ca-HAp failed to encounter any obvious improvement parameters. Nevertheless, on coating Galantamine by Ce/Ca-HAp all previous parameters retreated to near to normal levels. These results were simultaneously confirmed by pre-mentioned histological parameters.

CONCLUSION

According to the present findings multiple biomarkers would indeed improve diagnostic tools for the early detection of the onset of AD. Brain markers (Ach; Bcl-2; A β ; TPL; Chol; B-FABP; NO; MDA) together with brain antioxidants (GSH; SOD; CAT; CytoP450) may impose a progressive laboratory testing method besides well known imaging techniques. In addition, as Galantamine therapy may impose limited improvements thus the necessity for novel drug delivery systems to minimize dosing amounts, frequency of administration, and adverse side effects of drug while increasing its therapeutic efficacy is needed.

ACKNOWLEDGEMENT

The authors wish to acknowledge Dr. Ahmed Mohamed Abdellah, researcher at medical research centre, Ain Shams university hospitals where animals were housed subjected to ovariectomy prior to A β 13 for Alzheimer induction.

CONFLICTS OF INTEREST

No conflict of interest.

REFERENCES

- [1] Skoumalova A, Hort J. Blood markers of oxidative stress in Alzheimer's disease. *J Cell Mol Med* 2012; 16(10): 2291-2300. <http://dx.doi.org/10.1111/j.1582-4934.2012.01585.x>
- [2] Van Duijn CM, Clayton D, Chandra V, Fratiglioni L, Graves AB, Heyman A, Jorm AF, Kokmen E, Kondo K, Mortimer JA, *et al.* Familial aggregation of Alzheimer's disease and related disorders: a collaborative re-analysis of case-control studies. EURODEM Risk Factors Research Group. *Int J Epidemiol* 1991a; 20(2): 13-20. http://dx.doi.org/10.1093/ije/20.Supplement_2.S13
- [3] Van Duijn CM, Stijnen T, Hofman A. Risk factors for Alzheimer's disease: overview of the EURODEM collaborative re-analysis of case-control studies. EURODEM Risk Factors Research Group. *Int J Epidemiol* 1991b; 20(2): 4-12. http://dx.doi.org/10.1093/ije/20.Supplement_2.S4

- [4] Kivipelto M, Helkala E-L, Laakso MP, Hänninen T, Hallikainen M, Alhainen K, Soininen H, Tuomilehto J, Nissinen A. Midlife vascular risk factors and Alzheimer's disease in later life: longitudinal, population based study. *BMJ* 2001; 322: 1447-1451. <http://dx.doi.org/10.1136/bmj.322.7300.1447>
- [5] Doraiswamy PM, Krishen A, Stallone F, Martin WL, Potts NL, Metz A, DeVaugh-Geiss J. Cognitive performance on the Alzheimer's Disease Assessment Scale: effect of education. *Neurology* 1995; 45: 1980-1984. <http://dx.doi.org/10.1212/WNL.45.11.1980>
- [6] Galasko D, Abramson I, Corey-Bloom J, Thal LJ. Repeated exposure to the Mini-Mental State Examination and the Information-Memory-Concentration Test results in a practice effect in Alzheimer's disease. *Neurology* 1993; 43: 1559-1563. <http://dx.doi.org/10.1212/WNL.43.8.1559>
- [7] Manly JJ, Jacobs DM, Sano M, Bell K, Merchant CA, Small SA, Stern Y. Cognitive test performance among non dementia elderly African Americans and whites. *Neurology* 1998; 50: 1238-1245. <http://dx.doi.org/10.1212/WNL.50.5.1238>
- [8] Koivisto K. Population-based dementia screening program in the City of Kuopio, eastern Finland: evaluation of screening methods, prevalence of dementia and dementia subtypes. University of Kuopio, Department of Neurology Series of reports No 33. 1995.
- [9] DeCarli C, Haxby JV, Gillette JA, Teichberg D, Rapoport SI, Schapiro MB. Longitudinal changes in lateral ventricular volume in patients with dementia of the Alzheimer type. *Neurology* 1992; 42: 2029-2036. <http://dx.doi.org/10.1212/WNL.42.10.2029>
- [10] Talbot PR, Lloyd JJ, Snowden JS, Neary D, Testa HJ. A clinical role for 99mTc-HMPAO SPECT in the investigation of dementia? *J Neurol Neurosurg Psychiatry* 1998; 64: 306-313. <http://dx.doi.org/10.1136/jnnp.64.3.306>
- [11] Borroni B, DiLuca M, Cattabeni F, Padovani A. Advance on the diagnostic potential of biological markers in the early detection of Alzheimer Disease. *Neuroscience* 2004; 35(3): 232-245. <http://dx.doi.org/10.1002/nrc.20036>
- [12] Teunissen CE, de Vente J, Steinbusch HW, De Bruijn C. Biochemical markers related to Alzheimer's dementia in serum and cerebrospinal fluid. *Neurobiol Aging* 2002; 23: 485-508. [http://dx.doi.org/10.1016/S0197-4580\(01\)00328-1](http://dx.doi.org/10.1016/S0197-4580(01)00328-1)
- [13] Francis PT, Palmer AM, Snape M, Wilcock GK. The cholinergic hypothesis of Alzheimer's disease: a review of progress. *J. Neurol. Neurosurg. Psychiatry* 1999; 66(2): 137-147. <http://dx.doi.org/10.1136/jnnp.66.2.137>
- [14] Ramirez MJ. Differential involvement of 5-HT(1B/1D) and 5-HT6 receptors in cognitive and non-cognitive symptoms in Alzheimer's disease. *Neuropsychopharmacology* 2004; 29(2): 410-416. <http://dx.doi.org/10.1038/sj.npp.1300330>
- [15] Gil-Bea FJ, García-Alloza M, Marcos JB, Ramírez MJ. Evaluation of cholinergic markers in Alzheimer's disease and in a model of cholinergic deficit. *Neuroscience Letts* 2005; 375(1): 37-41. <http://dx.doi.org/10.1016/j.neulet.2004.10.062>
- [16] Tariot PN. Maintaining cognitive function in Alzheimer disease: how effective are current treatments. *Alzheimer Dis Assoc Disord* 2001; 15S: 26-33. <http://dx.doi.org/10.1097/00002093-200108001-00005>
- [17] Anglade P, Vyas S, Javoy-Agid F, *et al.* Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. *Histol Histopathol* 1997; 12: 25-31.
- [18] Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002; 297(5580): 353-356. <http://dx.doi.org/10.1126/science.1072994>
- [19] Steinerman JR, Irizarry M, Scarmeas N, *et al.* Distinct Pools of β -Amyloid in Alzheimer Disease—Affected Brain Arch Neurol 2008; 65(7): 906-912. <http://dx.doi.org/10.1001/archneur.65.7.906>
- [20] Carson SD. Tissue factor-initiated blood coagulation. *Prog Clin Pathol* 1984; 9: 1-14.
- [21] McComb RD, Miller KA, Carson SD. Tissue Factor Antigen in Senile Plaques of Alzheimer's Disease. *Am J Pathol* 1991; 139(3): 491-494.
- [22] Reed B, Villeneuve S, Mack W, DeCarli C, Chui H, Jagust W. Associations between serum cholesterol levels and cerebral amyloidosis. *JAMA Neurol* 2014; 71(2): 195-200. <http://dx.doi.org/10.1001/jamaneurol.2013.5390>
- [23] Hughes T. Cholesterol Metabolism in the Brain and Dementia. Doctoral Dissertation, University of Pittsburgh Pennsylvania, USA 2011.
- [24] Myers-Pane SC, Hubbel T, Pu L, Schnütgen F, Börschers T, Wood WG. Isolation and characterization of two fatty acid-binding proteins from mouse brain. *J Neurochem* 1996; 66: 1648-1656. <http://dx.doi.org/10.1046/j.1471-4159.1996.66041648.x>
- [25] Glatz JFC, Van der Vusse GJ. Cellular fatty acid-binding proteins: their function and physiological signification. *Prog Lipid Res* 1996; 3: 243-82. [http://dx.doi.org/10.1016/S0163-7827\(96\)00006-9](http://dx.doi.org/10.1016/S0163-7827(96)00006-9)
- [26] Glatz JFC, VanderVoort D, Hermens WT. Fatty acid binding protein as the earliest available plasma marker of acute myocardial injury. *J Clin Ligand Assay* 2002; 25: 167-77.
- [27] Pu L, Igbavboa U, Wood WG, Roths JB, Kier AB, Spener F. Expression of fatty acid binding protein is altered in aged mouse brain. *Mol Cell Biochem* 1999; 198: 69-78. <http://dx.doi.org/10.1023/A:1006946027619>
- [28] Shukla R. Nitric oxide in neurodegeneration. *Ann Neurosc* 2007; 14(1): 13-20. <http://dx.doi.org/10.5214/ans.0972.7531.2007.140104>
- [29] Aliev G, Palacios HH, Lipsitt AE, *et al.* Nitric oxide as an initiator of brain lesions during the development of Alzheimer disease. *Neurotoxicity Research* 2009; 16(3): 293-305. <http://dx.doi.org/10.1007/s12640-009-9066-5>
- [30] Aliev G, Li Y, Palacios HH, Obrenovich ME. Oxidative stress induced mitochondrial DNA deletion as a hallmark for the drug development in the context of the cerebrovascular diseases. *Recent Pat Cardiovasc Drug Discov* 2011; 6(3): 222-224. <http://dx.doi.org/10.2174/157489011797376942>
- [31] Moncada S, Erusalimsky JD. Does nitric oxide modulate mitochondrial energy generation and apoptosis? *Nat Rev Mol Cell Biol* 2002; 3(3): 214-220. <http://dx.doi.org/10.1038/nrm762>
- [32] Greilberger J, Koidl C, Greilberger M, *et al.* Malondialdehyde, carbonyl proteins and albumin-disulphide as useful oxidative markers in mild cognitive impairment and Alzheimer's disease. *Free Radic Res* 2008; 42: 633-638. <http://dx.doi.org/10.1080/10715760802255764>
- [33] Sinem F, Dildar K, Gö'khan E, *et al.* The serum protein and lipid oxidation marker levels in Alzheimer's disease and effects of cholinesterase inhibitors and antipsychotic drugs therapy. *Curr Alzheimer Res* 2010; 7: 463-469. <http://dx.doi.org/10.2174/156720510791383822>
- [34] Benzi G, Moretti A. Age- and peroxidative stress-related modifications of the cerebral enzymatic activities linked to mitochondria and the glutathione system. *Free Radic Biol Med* 1995; 19(1): 77-101. [http://dx.doi.org/10.1016/0891-5849\(94\)00244-E](http://dx.doi.org/10.1016/0891-5849(94)00244-E)

- [35] Padurariu M, Ciobica A, Hritcu L, Stoica B, Bild W, Stefanescu C. Changes of some oxidative stress markers in the serum of patients with mild cognitive impairment and Alzheimer's disease. *Neuroscience Lett* 2010; 469(1): 6-10. <http://dx.doi.org/10.1016/j.neulet.2009.11.033>
- [36] Kihara T, Sawada H, Nakamizo T, Kanki R, Yamashita H, Maelicka A, Shimohama S. Galanamine modulates nicotinic receptor and blocks Abeta-enhanced glutamate toxicity. *Biochem Biophys Res Commun* 2004; 325: 976-982. <http://dx.doi.org/10.1016/j.bbrc.2004.10.132>
- [37] Hwang J, Rodgers K, Oliver JC, Schlupe T. α -methylprednisolone conjugated cyclodextrin polymer based nanoparticles for rheumatoid arthritis therapy. *Int J Nanomedicine* 2008; 3(3): 359-372.
- [38] Riman RE, Suchanek WL, Byrappa K, Chen CW, Shuk P, Oakes CS. Solution synthesis of hydroxyapatite designer particulates. *Solid State Ionics* 2001; 151: 393-402. [http://dx.doi.org/10.1016/S0167-2738\(02\)00545-3](http://dx.doi.org/10.1016/S0167-2738(02)00545-3)
- [39] Estevez AY, Erlichman JS. Cerium Oxide Nanoparticles for the Treatment of Neurological Oxidative Stress Diseases. *Am Chem Soc* 2011; 1083: 255-288. <http://dx.doi.org/10.1021/bk-2011-1083.ch009>
- [40] Luo Y, Teng Z, Wang X, Wang Q. Development of carboxymethyl chitosan hydrogel beads in alcohol-aqueous binary solvent for nutrient delivery applications. *Food Hydrocolloids* 2012; 31: 332-339. <http://dx.doi.org/10.1016/j.foodhyd.2012.11.011>
- [41] Takuma K, Matsuo A, Himeno Y, *et al.* 17 beta-estradiol attenuates hippocampal neuronal loss and cognitive dysfunction induced by chronic restraint stress in ovariectomized rats. *Neuroscience* 2007; 146: 60-68. <http://dx.doi.org/10.1016/j.neuroscience.2007.01.017>
- [42] Krasovskii GN, Vasukovich LY, Chariev OG. Experimental study of biological effects of lead and aluminium following oral administration. *Environ Health Perspect* 1979; 30: 47-51.
- [43] Iliiev AI, Traykov VB, Mantchev GT, *et al.* A post-ischemic single administration of galantamine, a cholinesterase inhibitor, improves learning ability in rats. *J Pharm Pharmacol* 2000; 52: 1151-1156. <http://dx.doi.org/10.1211/0022357001774921>
- [44] Ellman GL, Courtney KD. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol* 1961; 7: 88-90. [http://dx.doi.org/10.1016/0006-2952\(61\)90145-9](http://dx.doi.org/10.1016/0006-2952(61)90145-9)
- [45] Barbareschi M, Veronese S, Leek R, Fox S, Bonzanini M, Girlando S, *et al.* Bcl-2 and P53 expression in node-negative breast carcinoma—study with long-term follow-up. *Hum Pathol* 1996; 27(11): 1149-1155. [http://dx.doi.org/10.1016/S0046-8177\(96\)90307-X](http://dx.doi.org/10.1016/S0046-8177(96)90307-X)
- [46] David EK, Claus UP, Edward HK. Modulation of amyloid B-protein clearance and Alzheimer's disease induced in rats. *J Clin Invest* 2000; 106: 1159-1166. <http://dx.doi.org/10.1172/JCI11013>
- [47] Bolhuis PA, Sylva-Steenland RMR, Tutuarima JA, Hische EAH. Comparison of the spectrophotometric determination and the two-stage coagulation assay of tissue factor activity. *Thrombosis Research* 1982; 27: 429-434. [http://dx.doi.org/10.1016/0049-3848\(82\)90060-3](http://dx.doi.org/10.1016/0049-3848(82)90060-3)
- [48] Sidel J, Haegele E, Wahlefeld A. Reagent for the enzymatic determination of serum total cholesterol with improved lipolytic efficiency. *Clin Chem* 1983; 29: 1075-1078.
- [49] Liu RZ, Mita R, Beaulieu M, Gao Z, Godbout R. Fatty acid binding proteins in brain development and disease. *Int J Dev Biol* 2010; 54(8-9): 1229-1239. <http://dx.doi.org/10.1387/ijdb.092976r1>
- [50] Nathan SB, Matthew BG. Methods to detect nitric oxide and its metabolites in biological samples. *Free Rad Biol & Med* 2007; 43: 645-657. <http://dx.doi.org/10.1016/j.freeradbiomed.2007.04.026>
- [51] Moore K, Roberts LJ. Measurement of lipid peroxidation. *Free Radic Res* 1998; 28: 659-671. <http://dx.doi.org/10.3109/10715769809065821>
- [52] Tietze F. Enzymic method for quantitative determination of nanogram amounts of total and oxidized glutathione: Applications to mammalian blood and other tissues. *Anal Biochem* 1969; 27: 502-522. [http://dx.doi.org/10.1016/0003-2697\(69\)90064-5](http://dx.doi.org/10.1016/0003-2697(69)90064-5)
- [53] Kuthan H, Haussmann HJ, Werringloer J. A spectrophotometric assay for superoxide dismutase activities in crude tissue fractions. *Biochem J* 1986; 237: 175-180. <http://dx.doi.org/10.1042/bj2370175>
- [54] Góth L. A simple method for determination of serum catalase activity and revision of reference range. *Clin Chim Acta* 1991; 196: 143-151. [http://dx.doi.org/10.1016/0009-8981\(91\)90067-M](http://dx.doi.org/10.1016/0009-8981(91)90067-M)
- [55] Schenkman JB. Cytochrome P450, "Handbook of Experimental Pharmacology", (Ed By Sinclair JF, Sinclair PR), 1993; 105: 259-277.
- [56] Harris HF. After Bruce Casselman WG. *Histopathological Technique*. Methuen and Co. Ltd 1959; (1900).
- [57] Querfurth HW, LaFerla FM. Alzheimer's disease. *N Engl J Med* 2010; 362: 329-344. <http://dx.doi.org/10.1056/NEJMra0909142>
- [58] Pratico D, Trojanowski JQ. Inflammatory hypotheses: novel mechanisms of Alzheimer's neurodegeneration and new therapeutic targets? *Neurobiol Aging* 2000; 21: 441-445.
- [59] Mahdy K, Shaker O, Wafay H, Nassar Y, Hassan H, Hussein A. Effect of some medicinal plant extracts on the oxidative stress status in Alzheimer's disease induced in rats. *Eur Rev Med Pharmacol Sci* 2012; 16(3): 31-42.
- [60] Yassin NA, El-Shenawy SM, Mahdy KA, Gouda NA, Marrie AFH, Farrag AH, Ibrahim BM. Effect of *Boswellia serrata* on Alzheimer's disease induced in rats. *JASMR* 2013; 8(1): 1-1170.
- [61] Vallés SL, Borrás C, Gambini J, Furriol J, Ortega A, Sastre J, Pallardo FV, Vina J. Oestradiol or genistein rescues neurons from amyloid beta-induced cell death by inhibiting activation of p38. *Aging Cell* 2008; 7(1): 112-118. <http://dx.doi.org/10.1111/j.1474-9726.2007.00356.x>
- [62] Wu Z, Du Y, Xue Hm, Wu Y, Zhou B. Aluminum induces neurodegeneration and its toxicity arises from increased iron accumulation and reactive oxygen species (ROS) production. *Neurobiol Aging* 2012; 33(1): 199-211. <http://dx.doi.org/10.1016/j.neurobiolaging.2010.06.018>
- [63] Terry AV, Buccafusco JJ. The cholinergic hypothesis of age and Alzheimer's disease related cognitive deficits: Recent challenges and their implications for novel drug development. *JPET* 2003; 306(3): 821-827. <http://dx.doi.org/10.1124/jpet.102.041616>
- [64] MacGibbon GA, Lawlor PA, Walton M. Expression of fos, jun, and krox family proteins in Alzheimer's disease. *Exp Neurol* 1997; 147(2): 316-332. <http://dx.doi.org/10.1006/exnr.1997.6600>
- [65] Hockenberry DM, Oltvai ZN, Yin X-M, Milliman CL, Korsmeyer SJ. Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell* 1993; 75: 241-251. [http://dx.doi.org/10.1016/0092-8674\(93\)80066-N](http://dx.doi.org/10.1016/0092-8674(93)80066-N)
- [66] Prehn JHM, Bindokas VP, Marcucelli CJ, Krajewski S, Reed JC, Miller RJ. Regulation of neuronal Bcl-2 protein expression and calcium homeostasis by transforming growth factor type β 1 confers wide-ranging protection on rat hippocampal neurons. *Proc Natl Acad Sci USA* 1994; 91(3): 1259-1261.
- [67] Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev* 2001; 81(2): 741-766.
- [68] Sadigh-Eteghad S, Saberमारouf B, Majdi A, Talebi M, Farhoudi M, Javad Mahmoudi J. Amyloid-Beta: A Crucial

- Factor in Alzheimer's Disease. *Med Princ Pract* 2015; 24: 1-10.
<http://dx.doi.org/10.1159/000369101>
- [69] Casserly I, Topol E. Convergence of atherosclerosis and Alzheimer's disease: inflammation, cholesterol, and misfolded proteins. *Lancet* 2004; 363: 1139-1146.
[http://dx.doi.org/10.1016/S0140-6736\(04\)15900-X](http://dx.doi.org/10.1016/S0140-6736(04)15900-X)
- [70] Eckert GP, Müller WE, Wood WG. Cholesterol-lowering Drugs and Alzheimer's Disease. *Future Lipidology* 2007; 2(4): 423-432.
<http://dx.doi.org/10.2217/17460875.2.4.423>
- [71] Simons M, Keller P, Dichgans J, Schulz JB. Cholesterol and Alzheimer's disease: is there a link? *Neurology* 2001; 57: 1089-1093.
<http://dx.doi.org/10.1212/WNL.57.6.1089>
- [72] Teunissen CE, Veerhuis R, De Vente J, Verhey RJF, Vreeling F, van Boxtel MPJ, Glatz JFC, Pelsers AL. Brain-specific fatty acid-binding protein is elevated in serum of patients with dementia-related diseases. *Eur J Neurol* 2011; 18(6): 865-871.
<http://dx.doi.org/10.1111/j.1468-1331.2010.03273.x>
- [73] Cho DH, Nakamura T, Fang J, Cieplak P, Godzik A, Gu Z, Lipton SA. S-Nitrosylation of Drp1 mediates β -Amyloid related mitochondrial fission and neuronal injury. *Science* 2009; 324: 102-105.
<http://dx.doi.org/10.1126/science.1171091>
- [74] Naditz A. Toxic Breakups: The Deadly Relationship Between Alzheimer's Disease and Nitric Oxide Gas. *Eukaryon* 2011; 7: 58-59.
- [75] Persson T, Popescu BO, Cedazo-Minguez A. Oxidative Stress in Alzheimer's Disease: Why Did Antioxidant Therapy Fail? *Oxid Med Cell Longev* 2014; 2014(2): 1-11.
<http://dx.doi.org/10.1155/2014/427318>
- [76] Khan RA, Khan MR, Sahreen S. Brain antioxidant markers, cognitive performance and acetylcholinesterase activity of rats: efficiency of *Sonchus asper*. *Behav Brain Funct* 2012; 8: 21-27.
<http://dx.doi.org/10.1186/1744-9081-8-21>

Received on 18-04-2016

Accepted on 08-06-2016

Published on 26-09-2016

DOI: <http://dx.doi.org/10.12970/2310-8231.2016.04.01.4>